Large Molecule Application of Volumetric Absorptive Microsampling for Single-Rodent PK Profiling of Exenatide Determined by LC-MRM

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OVERVIEW

PURPOSE

To demonstrate the applicability of Volumetric Absorptive Microsampling (VAMs) technology for the generation of a PK-profile derived from a single rodent for exenatide, a 4.2 kDa GLP-1 agonist biotherapeutic used in the treatment of type 2 diabetes.
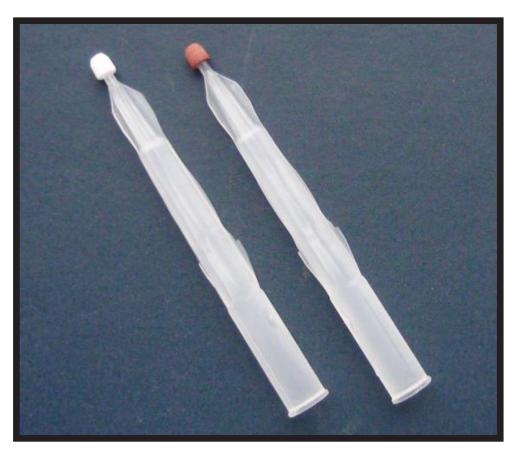
METHOD

Rat blood at each time-point was sampled onto a Mitra™ microsampling device (10 μL), dried for 24 hours at RT in the presence of desiccant, and extracted using Impact-Assisted Extraction in the presence of 70% acetonitrile. Data was acquired using a SCIEX Triple Quad 6500+ operated in positive ESI-MRM mode.
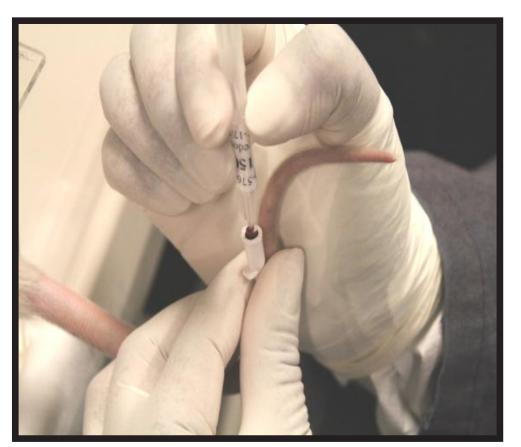
RESULTS

The VAMs method for exenatide demonstrated a linear range of 2.0 -1000 ng/mL with intra-assay precision and accuracy ≤ 8.8% and ≤ 2.6%, respectively. Average recovery was 75%, obtained without hematocrit effect. Individual PK profiles successfully derived from three subjects were in excellent agreement.

INTRODUCTION

Microsampling strategies in pre-clinical research allow consolidation of satellite TK and main study groups in an effort to replace, refine and reduce animal numbers. Additionally, toxicological effects can be correlated with exposure in the same individual. Capillary microsampling techniques circumvent the hematocrit (HCT) effect often reported for DBS analysis. However, processing is tedious and drugs exhibiting non-specific binding or requiring matrix stabilization are problematic. A recent alternative is Volumetric Absorptive Microsampling (VAMs), wherein an accurate volume of blood is absorbed onto a hydrophilic polymeric tip, thereby simplifying sample collection. The applicability of VAMs for the generation of a PK profile derived from a single rodent sampling only 10 μL of blood per time-point is examined for the biotherapeutic exenatide.

Figure 1. Neoteryx Mitra™ 10 μL microsampling tips used to collect blood from rat tail vein using an abbocath.

METHODS

PK STUDY IN SPRAGUE DAWLEY® RATS

- Exenatide dose: 210 μg subcutaneous
- 10 μl Mitra™ blood samples collected from tail vein
- Subjects: 3
- Total blood volume = 300 μL/rat (10 time-points collected in triplicate)

VAMS EXTRACTION PROCEDURE

Mitra™ Tip (10 μL), $^{13}C_9$^{15}N-exenatide (10 μL), 150 μL AcN:H$_2$O (70:30)

↓

Impact-Assisted Extraction in 70% ACN followed by centrifugation

↓

Aliquot 80 μL of supernatant, dilute with H$_2$O (100 μL)

LC-MS/MS DETECTION

- Shimadzu Nexera X2 UHPLC system
- Waters XBridge C$_{18}$ BEH Peptide column
- Gradient elution with 0.2% CH$_3$OOH in H$_2$O and AcN:MeOH (8:2)
- SCIEX Triple Quad API 6500+ operated in (+) ESI-MRM

Exenatide
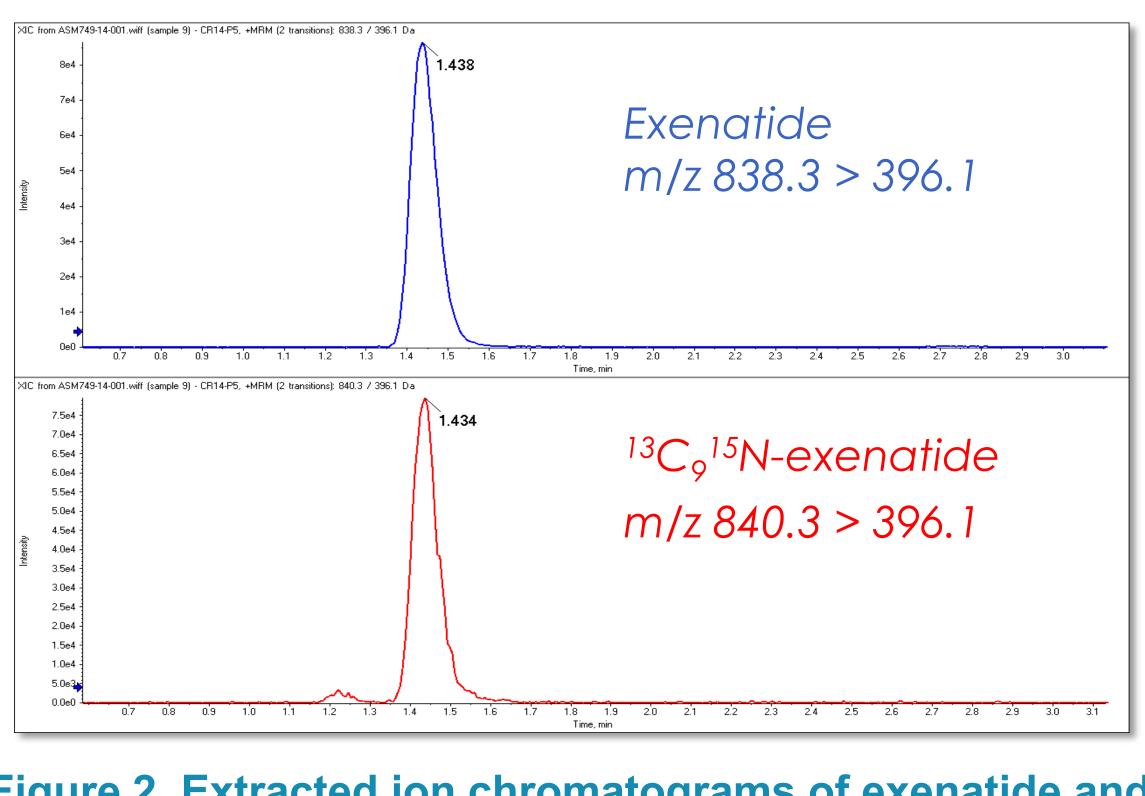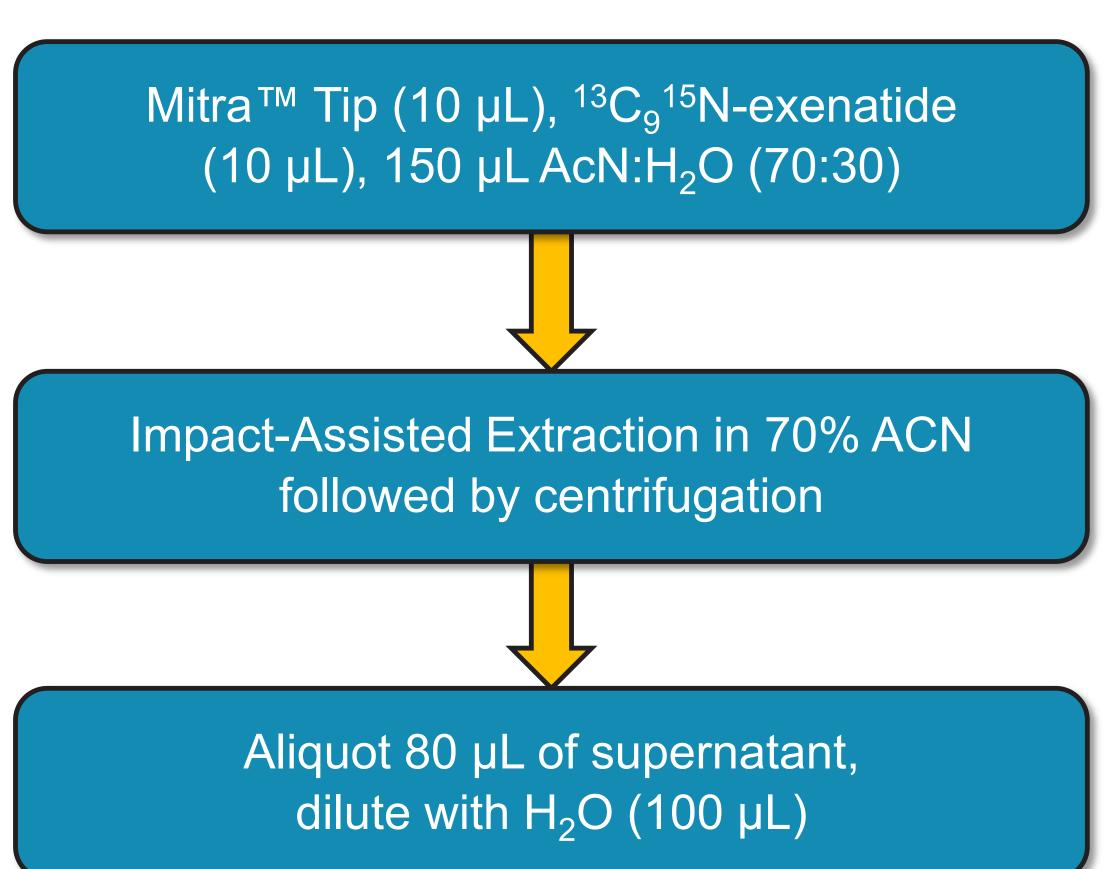m/z 838.3 > 396.1

$^{13}C_9$^{15}N-exenatide
m/z 840.3 > 396.1

Figure 2. Extracted ion chromatograms of exenatide and $^{13}C_9$^{15}N-exenatide (IS) with corresponding MRM transitions.

RESULTS

ADVANTAGES OF VAMS MICROSAMPLING
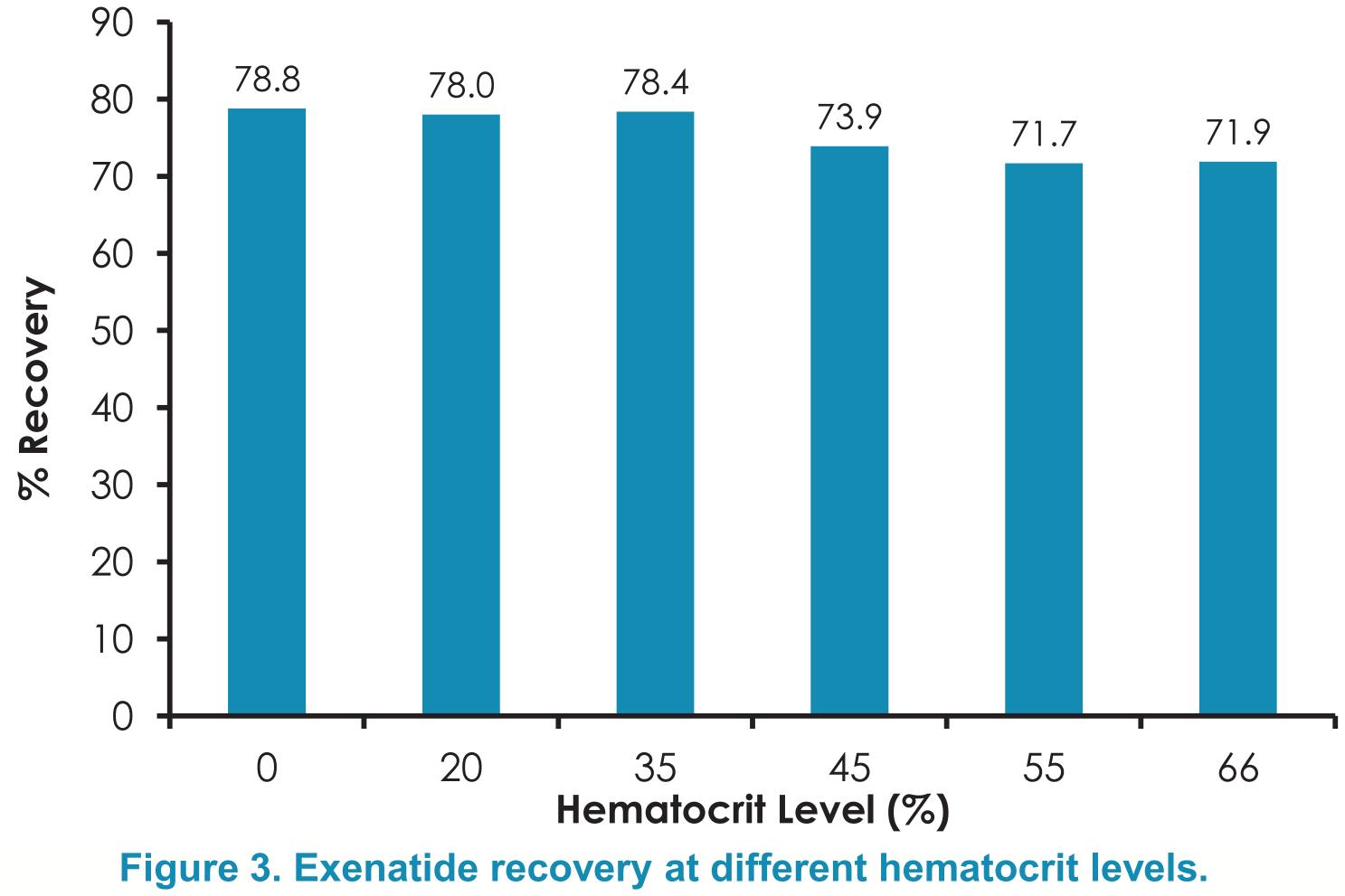
Sampling Volume vs. Blood Hematocrit

Precision and accuracy data for Mitra™ sampling volume, determined gravimetrically as a function of blood hematocrit level, revealed an absence of hematocrit effect as noted in Table 1 below.

Table 1. Hematocrit effect on blood sampling volume using VAMS.

Replicate	HCT = 0	HCT = 30	HCT = 44	HTC = 55	HTC = 66
Average Volume (μL)	10.56	10.40	10.58	10.69	10.74
N	12	12	12	12	12
% CV	2.7	3.0	3.1	3.3	3.5
Difference from HCT 44	-0.1%	-1.7%	0.0%	1.1%	1.6%

Effect of Hematocrit Level on Recovery

Exenatide at high QC concentration extracted from blood with hematocrit levels between 20% and 66% referenced against plasma (0% HCT) indicated minimal recovery bias between average blood yields and plasma. Further, recovery differences within the normal HCT range of 34% - 48% were largely insignificant (Figure 3).

Figure 3. Exenatide recovery at different hematocrit levels.

SENSITIVITY, LINEARITY, ACCURACY AND PRECISION

A major challenge in the analysis of biotherapeutics is low sensitivity due to multiple charging and variable fragmentation. For exenatide MS analysis (+5 charge-state monitored), sensitivity was notably improved using the SCIEX 6500+ when compared to the API 5500 (Figure 4).

API 6500+
API 5500
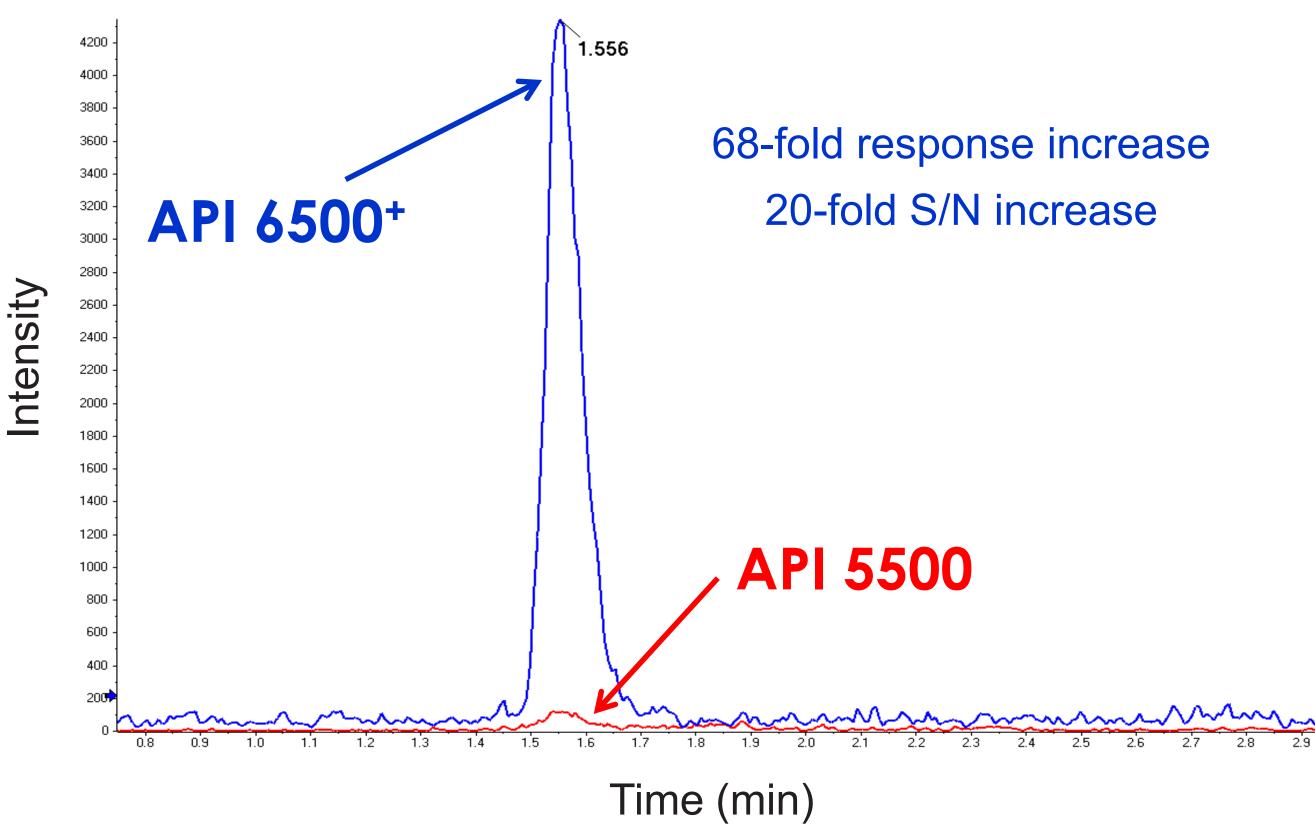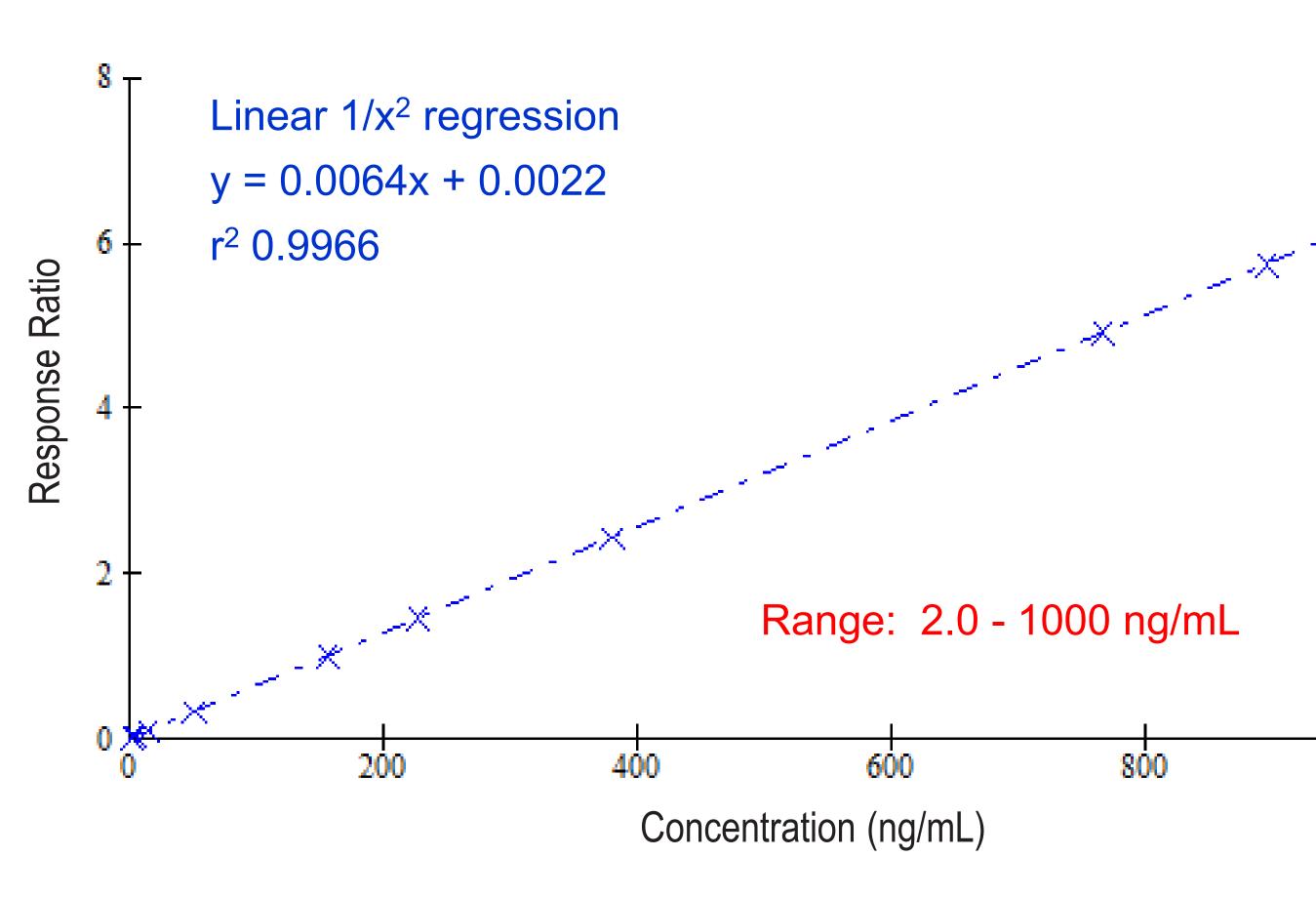68-fold response increase
20-fold S/N increase

Figure 4. Sensitivity comparison of SCIEX API 5500 vs. 6500+ for an extracted LOQ sample of exenatide (2.0 ng/mL, 540 fg on-column).

As illustrated in Figure 5, calibrants extracted from Mitra™ demonstrated excellent linearity and precision (r^2 = 0.9966) within a 500-fold analytical range. Selectivity, matrix effect and matrix factor all met acceptance criteria. Further, low and high QCs prepared in blood at HCT levels between 20 - 66% demonstrated back-calculated concentrations within ±15% when determined against calibrants prepared in blood at a HCT level of 44%.

Linear 1/x^2 regression
y = 0.0064x + 0.0022
r^2 0.9966
Range: 2.0 - 1000 ng/mL

Figure 5. Exenatide calibration curve extracted from Mitra™.

Extraction of QC samples in three different batches demonstrated excellent between-run precision and accuracy as highlighted in Table 2.

Table 2. Exenatide between-run precision and accuracy.

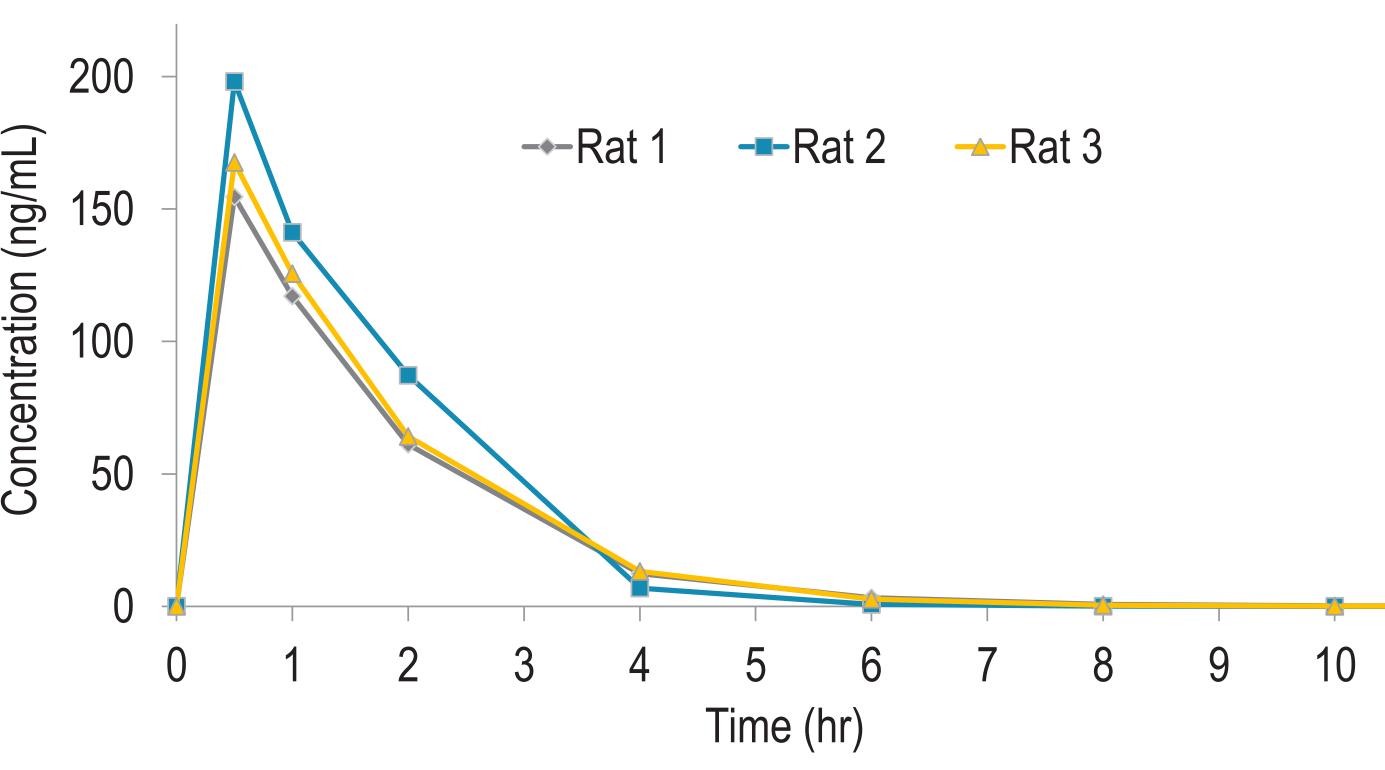| Stats	Concentration (ng/mL)			
	LOQ QC 2.00 ng/mL	QC1 6.00 ng/mL	QC2 200.00 ng/mL	QC3 750.00 ng/mL
Mean	2.05	6.06	204.89	769.32
S.D.	0.12	0.40	12.99	67.60
N	18	18	18	18
% C.V.	5.7	6.6	6.3	8.8
% Nominal	102.5	100.9	102.4	102.6

PK APPLICATION

Figure 6. PK profiles of exenatide in Sprague Dawley® rats (n = 3) following a single SC dose of 210 μg.

CONCLUSION

In this reported application of VAMs technology for the quantitation of a biotherapeutic, the technique demonstrated excellent linearity, precision and accuracy for exenatide extracted from rat blood whilst negating hematocrit effect. Further, sample collection using Mitra™ was significantly simplified when compared to alternative techniques such as DBS or capillary microsampling.